# Oxidative damage-related genes AKR1C3 and OGG1 modulate risks for lung cancer due to exposure to PAH-rich coal combustion emissions

Qing Lan<sup>1,7</sup>, Judy L.Mumford<sup>2</sup>, Min Shen<sup>1</sup>, David M.DeMarini<sup>2</sup>, Matthew R.Bonner<sup>1</sup>, Xingzhou He<sup>3</sup>, Meredith Yeager<sup>1</sup>, Robert Welch<sup>1</sup>, Stephen Chanock<sup>1</sup>, Linwei Tian<sup>4</sup>, Robert S.Chapman<sup>2</sup>, Tongzhang Zheng<sup>5</sup>, Phouthone Keohavong<sup>6</sup>, Neil Caporaso<sup>1</sup> and Nathaniel Rothman<sup>1</sup>

<sup>1</sup>Division of Cancer Epidemiology and Genetics, National Cancer Institute, NIH, DHHS, Bethesda, MD 20892, USA, <sup>2</sup>National Health and Environmental Effects Research Laboratory, United States Environmental Protection Agency, Research Triangle Park, NC 27711, USA, <sup>3</sup>Chinese Academy of Preventive Medicine, Beijing, China, <sup>4</sup>University of California, Berkeley, CA 94720, USA, <sup>5</sup>Yale School of Public Health, Yale University, New Haven, CT 06520, USA and <sup>6</sup>Department of Environmental and Occupational Health, University of Pittsburgh, Pittsburgh, PA 15260, USA

<sup>7</sup>To whom correspondence should be addressed Email: qingl@mail.nih.gov

Lung cancer rates among men and particularly among women, almost all of whom are non-smokers, in Xuan Wei County, China are among the highest in China and have been causally associated with exposure to indoor smoky coal emissions that contain very high levels of polycyclic aromatic hydrocarbons (PAHs). As such, this population provides a unique opportunity to study the pathogenesis of PAH-induced lung cancer that is not substantially influenced by the large number of other carcinogenic constituents of tobacco smoke. Aldo-keto reductases (AKRs) activate PAH dihydrodiols to yield their corresponding reactive and redox-active o-quinones, which can then generate reactive oxygen species that cause oxidative DNA damage. We therefore examined the association between single nucleotide polymorphisms (SNPs) in four genes (AKR1C3-Gln5His, NQO1-Pro187Ser, MnSOD-Val16Ala and OGG1-Ser326Cys) that play a role in the generation, prevention or repair of oxidative damage and lung cancer risk in a population-based, case-control study of 119 cases and 113 controls in Xuan Wei, China. The AKR1C3-Gln/Gln genotype was associated with a 1.84-fold [95% confidence interval (CI) = 0.98-3.45] increased risk and the combined OGG1-Cys/Cys and Ser/Cys genotypes were associated with a 1.93-fold (95% CI = 1.12-3.34) increased risk of lung cancer. Subgroup analysis revealed that the effects were particularly elevated among women who had relatively high cumulative exposure to smoky coal. SNPs in MnSOD and NQO1 were not associated with lung cancer risk. These results suggest that SNPs in the oxidative stress related-genes AKR1C3 and OGG1 may play a role in the pathogenesis of lung cancer in this population, particularly among heavily exposed women. However, due to the small sample size, additional studies are needed to

**Abbreviations:** AKR, aldo-keto reductases; PAH, polycyclic aromatic hydrocarbons; ROS, reactive oxygen species; SNP, single nucleotide polymorphisms.

evaluate these associations within Xuan Wei and other populations with substantial exposure to PAHs.

### Introduction

Residents in Xuan Wei, China are exposed to exceptionally high levels of polycyclic aromatic hydrocarbons (PAHs) from conception through adulthood via the smoky coal they burn for heating and cooking (1-5). Non-smoking women can inhale 10-times more PAH than a 20 cigarette/day active smoker, and air concentrations can approach levels experienced by workers on the top-side of coke ovens (5). Lung cancer rates in Xuan Wei County, China are among the highest in China and are similar for men and women (27.7 and 25.3 per 100 000 for men and women in the county, respectively) (5), despite the fact that almost all women are non-smokers. The lung cancer mortality rate in the three Xuan Wei communes where the most common smoky coal used has a particularly high polycyclic aromatic hydrocarbon (PAH) content was 118.0 and 125.6 per 100 000, for men and women, respectively, adjusted to the Chinese 1964 population, and 186.8 and 193.4 per 100 000 when adjusted to the US 1970 population.

Previous studies in this population have provided strong support that the lung cancer excess here is caused by smoky coal use among both men and women with only a modest contribution made by tobacco use among men (1,3–6). As such, the population in Xuan Wei provides a relatively unique opportunity to study the pathogenesis of PAH-induced lung cancer that is not influenced by the large number of other carcinogenic constituents of tobacco.

PAHs are activated to genotoxic intermediates through at least three primary pathways (Figure 1), all of which can lead to the production of G to T transversions (7). One of these pathways involves dihydrodiol dehydrogenases, which are members of the aldo-keto reductase (AKR) superfamily, and this pathway has been shown experimentally to produce PAH metabolites that form DNA adducts or reactive oxygen species (ROS) leading to oxidative DNA damage, such as 8-hydroxy-2'-deoxyguanosine (8-oxo-dG) (7). This superfamily includes AKR1A1, AKR1C1, AKR1C2, AKR1C3 and AKR1C4 (7-10). AKR1C3 is a particularly important enzyme in metabolizing potent trans-dihydrodiols containing more than two rings (7). Auto-oxidation of the intermediate catechols and/or subsequent redox cycling of the quinones can generate ROS that cause oxidative DNA damage (7,11,12).

Although single nucleotide polymorphisms (SNPs) in several genes in this pathway could influence the risk for lung cancer associated with cigarette smoke (13–15), which contains PAHs, SNPs in the *AKR* gene itself have not yet been examined for their influence on risk for cancer in an epidemiology study. Other metabolic enzymes that play a role in this process include *NQO1*, which reduces quinones back to catechols (16) and *MnSOD*, which can detoxify ROS in some

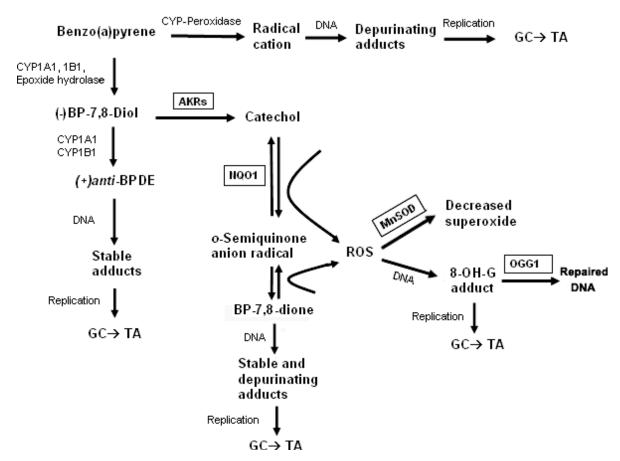


Fig. 1. Pathways for activation of benzo(a)pyrene, modified from Palackal et al. (1). The four genes whose SNPs were examined here are noted in boxes and span the entire pathway for the oxidative metabolism of PAHs.

instances (17). Multiple DNA repair mechanisms exist to avoid the effects of ROS and maintain genomic integrity (18,19). The most important DNA repair pathway of oxidative damage is base-excision repair (20) and includes the enzyme OGG1 (20–22).

Here we have evaluated the influence of a novel nonsynonymous SNP in *AKR1C3-Gln5His* on lung cancer risk, as well as SNPs in other genes in this pathway (*NQO1*, *SOD2* and *OGG1*), in a population-based, case-control study among residents of Xuan Wei County, China who have an extremely high PAH exposure due to the use of smoky coal in poorly vented homes (5). The results are considered in the context of previous reports on hot spot sites of PAH stable adducts in the *TP53* gene in lung tumors from tobacco smokers (23) and hot spots of G to T transversions in lung tumors from non-smoking women exposed to smoky coal in Xuan Wei (24).

## Materials and methods

This population-based, case-control study was described in detail elsewhere (3,25). Briefly, this study, which was carried out during the period from March 1995 to March 1996, contained a total of 122 newly diagnosed lung cancer cases. One control was selected for each lung cancer case, matching on sex, age (±2 years), village and type of fuel used currently for cooking and heating at home. As we described previously (3), the criteria for inclusion as a lung cancer case for this study were a histology or cytology confirmed (105 cases, 86.1%) or clinically diagnosed cases who died within a 1-year period (17 cases, 13.9%) (3). Information about demographic characteristics, lifetime consumption of different types of coal, tobacco smoking, family history of lung cancer and personal medical history were obtained from a standardized closed-question form of questionnaire. DNA was extracted from sputum samples

using phenol-chloroform extraction (26). Genotyping was carried out by real-time PCR on an ABI 7900HT sequence detection system as described on the SNP500 website (27). Of the 122 cases and 122 controls, DNA was successfully extracted from 119 cases and 113 controls. For human subject protection, this study was conducted according to the recommendations of the World Medical Association Declaration of Helsinki. The research protocol was approved by a US EPA Human Subjects Research Review Official for international research projects, and informed consent was obtained from all subjects in this study.

Genotype data were analyzed with the most common genotype as the referent, with the exception of AKR1C3, in which it is plausible that the most common variant could be associated with the greatest production of ROS. Genotype data for the four SNPs were not significantly correlated with each other or with pack-years of smoking, smoky coal use or the GSTM1 null genotype, which we have shown previously was associated with increased lung cancer risk in this population (3). The test for Hardy-Weinberg equilibrium among the controls was conducted using observed genotype frequencies and a  $\chi^2$  test with one degree of freedom. Unmatched unconditional logistic regression was used to estimate the odds ratios (ORs) and 95% confidence intervals (CIs), adjusting for age, sex, and pack-years of tobacco use, to test the association between lung cancer risk and genotypes and other risk factors so that all the genotype data could be used. Crude ORs were essentially identical to adjusted ORs, with the exception of the AKR1C3 association among women with heavy smoky coal use, which is presented in a footnote in Table III. Further adjustment for village and fuel type produced similar results in almost all models; however, in a few instances the ORs were substantially inflated, so these variables were not adjusted for in the analyses presented here. Data were analyzed using Statistical Analysis Software, version 8.02 (SAS Institute, 1996).

The OR for smoky coal use was estimated for the mean amount of lifetime cumulative tons of smoky coal, based on the distribution in the controls. In addition, smoky coal use was modeled as a linear relationship between the total amount of smoky coal used and the log odds of lung cancer. Gene–environment interactions were tested on a multiplicative scale by adding product terms into a logistic regression model.

### **Results**

Table I shows the distribution of characteristics for both cases and controls. Among cases and controls, age, sex and type of fuel source were comparable. Approximately 34% of the cases and controls were female. Ethnicity, education level, household income and dwelling type were comparable for cases and controls (not shown). About 93% of the males were tobacco smokers, whereas only one female smoked. Among men, smoking for >25 pack-years was associated with a 1.69-fold (95% CI = 0.82-3.49) increased risk of lung cancer. This relatively weak smoking effect was consistent with previous studies in Xuan Wei (1,3,4,6) and may have been due to the use of water pipes for smoking. Cases were more likely to be heavy smoky coal users than controls. Compared with subjects who used <130 tons of smoky coal during their lifetime, heavy smoky coal users (those who used >130 tons) had a 2.27-fold (95% CI = 1.25-4.10) increased risk of lung cancer. Among women, the risk was 6.11 (95% CI = 1.92-19.43) for heavy smoky coal use, and among men the risk was 1.49 (95% CI = 0.73 - 3.04).

Table I. Distribution of subject characteristics in lung cancer patients and controls

	Case <i>n</i> (%)	Control n (%)	P value <sup>a</sup>
Age			
< 55	50 (42)	46 (41)	
≥55	69 (58)	67 (59)	0.84
Sex			
Male	78 (66)	73 (65)	
Female	41 (34)	40 (35)	0.88
Smoking <sup>b</sup>			
No	5 (6)	5 (7)	
Yes	73 (94)	68 (93)	0.91
Smoky coal use			
< 130	50 (42)	67 (59)	
≥130	69 (58)	46 (41)	0.008

 $<sup>^{</sup>a}P$  based on  $\chi^{2}$  test.

Table II presents the association of genotypes with lung cancer risk. All genotype distributions were in Hardy-Weinberg equilibrium. We observed an increased lung cancer risk for the AKR1C3 and OGG1 genotypes, but not for the NQO1 or SOD2 genotypes (Table II). We further stratified the analysis of each genotype on lung cancer risk by both sex and smoky coal use in an attempt to separate the effects of each (Table III). There was a consistent pattern in that the impact of both the AKR1C3 and OGG1 genotypes on lung cancer risk was particularly pronounced among women exposed to >130 tons of smoky coal. The test for interaction between sex, smoky coal use as a dichotomous or continuous variable and each genotype was not, however, significant. Effects were similar among smokers with >25 pack-years of tobacco use and smokers with <25 pack-years, and tests for interaction with pack-year category or with pack-years as a continuous variable were not significant.

#### Discussion

The AKR superfamily, which includes AKR1C3, has been shown to oxidize PAHs to catechol, which can then produce a-dione that can form DNA adducts and/or ROS, which can cause oxidative damage to DNA (Figure 1). The AKR1C3-Gln5His SNP produces a glutamine-to-histidine amino acid change, but the impact on enzymatic activity and whether it is in linkage disequilibrium with other potentially functional variants is not known. Our results are consistent with the hypothesis that alterations in the relative contribution of AKR1C3 to the metabolism of PAHs could impact lung cancer risk. To our knowledge, this is the first report of a molecular epidemiological study that has suggested that genetic variation in AKR1C3 could be associated with risk for lung cancer. Comprehensive evaluation of additional SNPs in this gene are needed to estimate the importance of common haplotypes; moreover, additional studies of other AKR superfamily members are needed to characterize further the role of perturbations of these genes in the pathogenesis of PAH-associated lung cancer.

Table II. AKR1C3-Gln5His, NQO1-Pro187Ser, MnSOD2-Val16Ala and OGG1-Ser326Cys polymorphisms and lung cancer risk

Genotype	dbSNP ID	Case <i>n</i> (%)	Control n (%)	OR <sup>a</sup> (95% CI)	P value
AKR1C3	rs12529				
His/His		1 (0.86)	1 (0.89)	_	
His/Gln		21 (18.10)	32 (28.57)	_	
His/His + His/Gln		22 (18.97)	33 (29.46)	1.0	
Gln/Gln		94 (81.03)	79 (70.54)	1.84 (0.98-3.45)	0.06
NQO1	rs1800566	, , , ,	· · ·	, , , ,	
Pro/Pro		37 (31.09)	32 (29.36)	1.0	
Pro/Ser		57 (47.90)	54 (49.54)	0.90 (0.49-1.66)	0.75
Ser/Ser		25 (21.01)	23 (21.10)	0.90 (0.43-1.90)	0.79
Pro/Ser + Ser/Ser		82 (68.91)	77 (70.64)	0.91 (0.51-1.62)	0.73
SOD2	rs1799725				
Val/Val		93 (78.15)	81 (72.32)	1.0	
Val/Ala		23 (19.33)	30 (26.79)	0.67 (0.36-1.25)	0.20
Ala/Ala		3 (2.52)	1 (0.89)	_ ` `	
Val/Ala + Ala/Ala		26 (21.85)	31 (27.68)	0.73 (0.40-1.33)	0.30
OGG1	rs1052133				
Ser/Ser		37 (31.36)	51 (46.79)	1.0	
Ser/Cys		61 (51.69)	43 (39.45)	1.96 (1.10-3.57)	0.02
Cys/Cys		20 (16.95)	15 (13.76)	1.85 (0.83-4.11)	0.13
Ser/Cys + Cys/Cys		81 (68.64)	58 (53.21)	1.93 (1.12–3.34)	0.02

<sup>&</sup>lt;sup>a</sup>ORs and 95% CIs obtained by logistic regression analysis adjusted for age, sex, pack-years of smoking.

bMales only.

Table III. AKR1C3-Gln5His and OGG1-Ser326Cys polymorphisms and lung cancer risk stratified by smoky coal use and sex

Genotype	Smoky coal use without a chimney (tons)							
	<130 tons				≥130 tons			
	Case n (%)	Control n (%)	OR <sup>a</sup> (95%CI)	P value	Case n (%)	Control n (%)	OR <sup>a</sup> (95% CI)	P value
Female								
AKR1C3								
His/His + His/Gln	4 (33.33)	8 (33.33)	1.0		5 (17.24)	10 (66.67)	1.0	
Gln/Gln	8 (66.67)	16 (66.67)	1.00 (0.19-5.80) <sup>b</sup>	1.00	24 (82.76)	5 (33.33)	12.93 (2.20–107.82) <sup>bc</sup>	0.0018
OGG1								
Ser/Ser	5 (41.67)	11 (47.83)			4 (13.79)	8 (53.33)		
Ser/Cys + Cys/Cys	7 (58.33)	12 (52.17)	1.29 (0.31-5.28)	0.73	25 (86.21)	7 (46.67)	5.67 (1.07-34.16) <sup>b</sup>	0.04
Male								
AKR1C3								
His/His + His/Gln	8 (21.62)	11 (25.58)	1.0		5 (13.16)	4 (13.33)	1.0	
Gln/Gln	29 (78.38)	32 (74.42)	1.49 (0.50-4.43)	0.48	33 (86.84)	26 (86.67)	0.94 (0.17-4.95) <sup>b</sup>	1.00
OGG1	` ′	, ,	, ,		` /	, ,	`	
Ser/Ser	15 (40.54)	18 (43.90)			13 (32.50)	14 (46.67)		
Ser/Cys + Cys/Cys	22 (59.46)	23 (56.10)	1.14 (0.45-2.90)	0.78	27 (67.50)	16 (53.33)	2.04 (0.74-5.60)	0.17

<sup>&</sup>lt;sup>a</sup>ORs and 95% CIs obtained by logistic regression analysis adjusted for age, pack-year of smoking.

Consistent with the role of the AKR pathway in non-smokers with heavy smoky coal use is our finding that subjects with OGG1-Cys/Cys and Ser/Cys alleles were also at increased risk for lung cancer. Some (28,29) but not all (30) studies have provided evidence that this variant is associated with lung cancer risk, and there is evidence that it results in decreased DNA repair activity (13,31). OGG1 is part of the base-excision DNA repair system and removes oxidatively damaged bases, such as 8-oxo-G. Such damage would be expected as a consequence of metabolism of PAHs through the AKR pathway. In this regard, a recent study in mammalian cells found that OGG1 suppressed G to T transversions due to 8-oxo-G, but not those due to benzo[a]pyrene diol-epoxide adducts (32). Thus, we have considered the influence of this pathway on the mutation spectrum in smoky coal-associated lung tumors (24).

We identified previously three hot spots for mutations (G to T transversions) in the TP53 gene in lung tumors of nonsmoking women in Xuan Wei exposed to smoky coal emissions (24). Two of these mutational hot spots are at codons 154 and 273, which are also hot spots for PAH adduction (stable adducts) (23). We therefore postulated that stable PAH adducts, most likely produced through the CYP pathway (Figure 1) could be responsible for the G to T transversion at each site (24). However, the third mutational hot spot is at codon 249, which is not a hot spot for stable PAH adduction (23,24). Our present result leads us to suggest that G to T transversions at this site (codon 249) may be due to oxidative DNA damage resulting from the metabolism of PAHs in the smoky coal emissions via the AKR pathway. Such a pathway could produce unstable (depurinating) adducts via the production of the PAH-quinone, resulting in apurinic sites, or ROS that could result in oxidative damage, such as 8-oxo-G. Either type of damage, if not repaired, can be processed by the cell into G to T transversions (7,8,10). Thus, we propose that the mutation spectrum in smoky coal-associated tumors reflects PAH damage due both to stable adducts as well as to unstable adducts and/or oxidative base damage due to ROS production.

The *Pro187Ser* variant of *NQO1*, which is an oxidoreductase enzyme that plays a role in the reactions described in Figure 1, results in decreased enzyme activity (15,16); however, it was

not associated with lung cancer risk in our study. Overall, there have been both positive and null reports of its association with lung cancer (15,16,33). Similarly, the *Val16Ala* SNP in *SOD2*, which has been predicted to result in higher enzymatic activity (34), was not associated with increased risk in this study. There has been a positive association of this SNP with lung cancer risk in a Caucasian population but not in a Taiwanese population (14,33).

Our study has several strengths. It is a population-based, case-control study with high participation rates in a unique semi-mountainous rural area of China where environmental PAH exposure is 10-times higher than that received by a 1pack/day smoker and can approach concentrations experienced by coke-oven workers. This contrasts with almost all other molecular epidemiology studies of lung cancer reported previously in the literature. Further, our series of lung cancer cases is unique in that smoky coal exposure appears as the primary etiological factor, in contrast to tobacco in almost all other populations studied. This is particularly so for women in Xuan Wei, whose lung cancer incidence is the highest in China (5). As a consequence, this study population provides a unique group to explore the influence of variation in key genes that mediate the effects of PAHs on lung cancer risk. Finally, the pattern of particularly elevated genetic risks among women who were heavy smoky coal users is consistent with the effects of the GSTM1 null genotype on lung cancer risk in this population (unpublished data).

The primary limitation of our study is its small sample size and consequently low power. As pointed out recently by Wacholder *et al.* (35), a statistically significant association in a small study has an increased likelihood of being a false positive finding, particularly if the risk factor has a low prior probability. In addition, some investigators are concerned about multiple comparisons increasing the probability of detecting statistically significant yet false positive findings. We evaluated the overall effects of four SNPs and carried out a number of subgroup analyses. The observed low *P*-value of the association between the *AKR1C3-Gln/Gln* genotype and lung cancer in heavily exposed women would meet the standard of statistical significance that would be required

<sup>&</sup>lt;sup>b</sup>Exact test.

<sup>&</sup>lt;sup>c</sup>Crude OR = 9.60 (2.27-40.62), P = 0.0021.

by procedures that attempt to correct for multiple comparisons. Clearly, however, the findings from this study need to be considered preliminary. A new, substantially larger case-control study of lung cancer among non-smoking women is being planned among this relatively unique population and will provide an opportunity to replicate and extend these findings. Finally, more extensive analysis of *AKR1C3* is needed to comprehensively assess the contribution of genetic variation across the gene (36) to lung cancer risk and to study the impact of genetic variation on enzyme activity.

#### Acknowledgements

The authors are grateful to Dongyun Yang and Chaofu Huang for their assistance with the data. We thank R.Julian Preston, Stephen Nesnow, Russell Owen, Daniel Shaughnessy and Ron Rogers for their helpful comments on the manuscript. This manuscript has been reviewed by the National Health and Environmental Effects Research Laboratory, United States Environmental Protection Agency and approved for publication. Approval does not signify that the contents necessarily reflect the views and policies of the Agency, nor does mention of trade names or commercial products constitute endorsement or recommendation for use. P.K. was supported by American Cancer Society grant #RSG-99-161-04-CNE.

### References

- 1. Chapman,R.S., Mumford,J.L., Harris,D.B., He,Z.Z., Jiang,W.Z. and Yang,R.D. (1988) The epidemiology of lung cancer in Xuan Wei, China: current progress, issues, and research strategies. *Arch. Environ. Health*, **43**, 180–185.
- He,X.Z.a.Y.R.D. (1994) Lung Cancer and Indoor Air Pollution from Coal Burning. Yuan Nan Science and Technology Publishing House.
- 3. Lan,Q., He,X., Costa,D.J., Tian,L., Rothman,N., Hu,G. and Mumford,J.L. (2000) Indoor coal combustion emissions, *GSTM1* and *GSTT1* genotypes, and lung cancer risk: a case-control study in Xuan Wei, China. *Cancer Epidemiol. Biomarkers Prev.*, **9**, 605–608.
- Lan,Q., Chapman,R.S., Schreinemachers,D.M., Tian,L. and He,X. (2002) Household stove improvement and risk of lung cancer in Xuanwei, China. J. Natl Cancer Inst., 94, 826–835.
- Mumford,J.L., He,X.Z., Chapman,R.S., Cao,S.R., Harris,D.B., Li,X.M., Xian,Y.L., Jiang,W.Z., Xu,C.W. and Chuang,J.C. (1987) Lung cancer and indoor air pollution in Xuan Wei, China. *Science*, 235, 217–220.
- Liu, Z.Y., He, X.Z. and Chapman, R.S. (1991) Smoking and other risk factors for lung cancer in Xuanwei, China. Int. J. Epidemiol., 20, 26–31.
- 7. Palackal, N.T., Lee, S.H., Harvey, R.G., Blair, I.A. and Penning, T.M. (2002) Activation of polycyclic aromatic hydrocarbon trans-dihydrodiol proximate carcinogens by human aldo-keto reductase (AKR1C) enzymes and their functional overexpression in human lung carcinoma (A549) cells. *J. Biol. Chem.*, 277, 24799–24808.
- Palackal, N.T., Burczynski, M.E., Harvey, R.G. and Penning, T.M. (2001) Metabolic activation of polycyclic aromatic hydrocarbon trans-dihydrodiols by ubiquitously expressed aldehyde reductase (AKR1A1). *Chem. Biol. Interact.*, 130–132, 815–824.
- Penning, T.M., Burczynski, M.E., Jez, J.M., Hung, C.F., Lin, H.K., Ma, H., Moore, M., Palackal, N. and Ratnam, K. (2000) Human 3alpha-hydroxysteroid dehydrogenase isoforms (AKR1C1-AKR1C4) of the aldo-keto reductase superfamily: functional plasticity and tissue distribution reveals roles in the inactivation and formation of male and female sex hormones. *Biochem. J.*, 351, 67-77.
- Yu,D., Kazanietz,M.G., Harvey,R.G. and Penning,T.M. (2002) Polycyclic aromatic hydrocarbon o-quinones inhibit the activity of the catalytic fragment of protein kinase C. *Biochemistry*, 41, 11888–11894.
- 11. Flowers, L., Bleczinski, W.F., Burczynski, M.E., Harvey, R.G. and Penning, T.M. (1996) Disposition and biological activity of benzo[a]pyrene-7,8-dione. A genotoxic metabolite generated by dihydrodiol dehydrogenase. *Biochemistry*, 35, 13664–13672.
- 12. Flowers, L., Ohnishi, S.T. and Penning, T.M. (1997) DNA strand scission by polycyclic aromatic hydrocarbon o-quinones: role of reactive oxygen species, Cu(II)/Cu(I) redox cycling, and o-semiquinone anion radicals. *Biochemistry*, **36**, 8640–8648.
- Goode, E.L., Ulrich, C.M. and Potter, J.D. (2002) Polymorphisms in DNA repair genes and associations with cancer risk. *Cancer Epidemiol. Biomarkers Prev.*, 11, 1513–1530.

- 14. Wang, L.I., Miller, D.P., Sai, Y., Liu, G., Su, L., Wain, J.C., Lynch, T.J. and Christiani, D.C. (2001) Manganese superoxide dismutase alanine-to-valine polymorphism at codon 16 and lung cancer risk. J. Natl Cancer Inst., 93, 1818–1821.
- 15. Wiencke, J.K., Spitz, M.R., McMillan, A. and Kelsey, K.T. (1997) Lung cancer in Mexican-Americans and African-Americans is associated with the wild-type genotype of the NAD(P)H: quinone oxidoreductase polymorphism. *Cancer Epidemiol. Biomarkers Prev.*, 6, 87–92.
- 16. Sunaga, N., Kohno, T., Yanagitani, N., Sugimura, H., Kunitoh, H., Tamura, T., Takei, Y., Tsuchiya, S., Saito, R. and Yokota, J. (2002) Contribution of the *NQO1* and *GSTT1* polymorphisms to lung adenocarcinoma susceptibility. *Cancer Epidemiol. Biomarkers Prev.*, 11, 730–738.
- 17. Kinnula, V.L. and Crapo, J.D. (2003) Superoxide dismutases in the lung and human lung diseases. *Am. J. Respir. Crit. Care Med.*, **167**, 1600–1619.
- 18. Caporaso, N. (2003) The molecular epidemiology of oxidative damage to DNA and cancer. *J. Natl Cancer Inst.*, **95**, 1263–1265.
- Wei,Q., Cheng,L., Hong,W.K. and Spitz,M.R. (1996) Reduced DNA repair capacity in lung cancer patients. *Cancer Res.*, 56, 4103–4107.
- Slupphaug, G., Kavli, B. and Krokan, H.E. (2003) The interacting pathways for prevention and repair of oxidative DNA damage. *Mutat. Res.*, 531, 231–251.
- Shinmura, K., Kasai, H., Sasaki, A., Sugimura, H. and Yokota, J. (1997)
  8-Hydroxyguanine (7,8-dihydro-8-oxoguanine) DNA glycosylase and AP lyase activities of hOGG1 protein and their substrate specificity. *Mutat. Res.*, 385, 75–82.
- 22. Shinmura, K. and Yokota, J. (2001) The OGG1 gene encodes a repair enzyme for oxidatively damaged DNA and is involved in human carcinogenesis. *Antioxid. Redox. Signal.*, 3, 597–609.
- 23. Smith, L.E., Denissenko, M.F., Bennett, W.P., Li, H., Amin, S., Tang, M. and Pfeifer, G.P. (2000) Targeting of lung cancer mutational hotspots by polycyclic aromatic hydrocarbons. *J. Natl Cancer Inst.*, **92**, 803–811.
- 24. DeMarini, D.M., Landi, S., Tian, D. *et al.* (2001) Lung tumor KRAS and TP53 mutations in nonsmokers reflect exposure to PAH-rich coal combustion emissions. *Cancer Res.*, **61**, 6679–6681.
- 25. Lan, Q., Feng, Z., Tian, D., He, X., Rothman, N., Tian, L., Lu, X., Terry, M.B. and Mumford, J.L. (2001) p53 gene expression in relation to indoor exposure to unvented coal smoke in Xuan Wei, China. J. Occup. Environ. Med., 43, 226–230.
- 26. Garcia-Closas, M., Egan, K.M., Abruzzo, J. et al. (2001) Collection of genomic DNA from adults in epidemiological studies by buccal cytobrush and mouthwash. Cancer Epidemiol. Biomarkers Prev., 10, 687–696.
- 27. Packer, B.R., Yeager, M., Staats, B. *et al.* (2004) SNP500Cancer: a public resource for sequence validation and assay development for genetic variation in candidate genes. *Nucleic Acids Res.*, **32** (Database issue), D528–D532.
- 28. Sugimura, H., Kohno, T., Wakai, K. et al. (1999) hOGG1 Ser326Cys polymorphism and lung cancer susceptibility. Cancer Epidemiol. Biomarkers Prev., 8, 669–674.
- Le Marchand, L., Donlon, T., Lum-Jones, A., Seifried, A. and Wilkens, L.R. (2002) Association of the hOGG1 Ser326Cys polymorphism with lung cancer risk. *Cancer Epidemiol. Biomarkers Prev.*, 11, 409–412.
- 30. Wikman, H., Risch, A., Klimek, F., Schmezer, P., Spiegelhalder, B., Dienemann, H., Kayser, K., Schulz, V., Drings, P. and Bartsch, H. (2000) hOGG1 polymorphism and loss of heterozygosity (LOH): significance for lung cancer susceptibility in a caucasian population. Int. J. Cancer, 88, 932–937.
- 31. Kohno, T., Shinmura, K., Tosaka, M., Tani, M., Kim, S.R., Sugimura, H., Nohmi, T., Kasai, H. and Yokota, J. (1998) Genetic polymorphisms and alternative splicing of the *hOGG1* gene, that is involved in the repair of 8-hydroxyguanine in damaged DNA. *Oncogene*, 16, 3219–3225.
- 32. Yamane, A., Shinmura, K., Sunaga, N. *et al.* (2003) Suppressive activities of OGG1 and MYH proteins against G:C to T:A mutations caused by 8-hydroxyguanine but not by benzo[a]pyrene diol epoxide in human cells *in vivo. Carcinogenesis*, **24**, 1031–1037.
- 33. Lin, P., Hsueh, Y.M., Ko, J.L., Liang, Y.F., Tsai, K.J. and Chen, C.Y. (2003) Analysis of NQO1, GSTP1, and MnSOD genetic polymorphisms on lung cancer risk in Taiwan. *Lung Cancer*, **40**, 123–129.
- 34. Rosenblum, W.I., Nishimura, H. and Nelson, G.H. (1992) L-NMMA in brain microcirculation of mice is inhibited by blockade of cyclooxygenase and by superoxide dismutase. *Am. J. Physiol.*, **262**, H1343–H1349.
- 35. Wacholder, S., Chanock, S., Garcia-Closas, M., El Ghormli, L. and Rothman, N. (2004) Assessing the probability that a positive report is false: an approach for molecular epidemiology studies. *J. Natl Cancer Inst.*, **96**, 434–442.
- 36. Bader, J.S. (2001) The relative power of SNPs and haplotype as genetic markers for association tests. *Pharmacogenomics*, **2**, 11–24.

Received April 14, 2004; revised June 21, 2004; accepted July 18, 2004